

REVIEW ARTICLE

The many faces of p38 mitogen-activated protein kinase in progenitor/stem cell differentiation

Feride OEZTUERK-WINDER and Juan-Jose VENTURA¹

CSCR (Wellcome Trust Centre for Stem Cell Research), Tennis Court Road, Cambridge CB2 1QR, U.K.

Regulation of stem cells is essential for development and adult tissue homoeostasis. The proper control of stem cell self-renewal and differentiation maintains organ physiology, and disruption of such a balance results in disease. There are many mechanisms that have been established as stem cell regulators, such as Wnt or Notch signals. However, the intracellular mechanisms that mediate and integrate these signals are not well understood. A new intracellular pathway that has been reported to be involved in the regulation of many stem cell types is that of p38 MAPK (mitogen-activated protein kinase). In particular, p38 α is essential for the proper differentiation of many haematopoietic, mesenchymal and epithelial stem/progenitor cells. Many reports

have shown that disruption of this kinase pathway has pathological consequences in many organs. Understanding the extracellular cues and downstream targets of $p38\alpha$ in stem cell regulation may help to tackle some of the pathologies associated with improper differentiation and regulation of stem cell function. In the present review we present a vision of the current knowledge on the roles of the $p38\alpha$ signal as a regulator of stem/progenitor cells in different tissues in physiology and disease.

Key words: cancer, cell specification, cytokine signalling, development, mitogen-activated protein kinase (MAPK), self-renewal.

INTRODUCTION

Embryonic development and maintenance of adult tissue homoeostasis are physiological processes where stem cells, which are characterized by their ability to self-renew and perpetuate themselves, generate differentiated cell types or replenish functional tissue. Furthermore, they have the potential to produce differentiated daughter cells that will eventually become specialized embryonic or adult cells.

Many signalling pathway components are well known as key players in stem cell differentiation and self-renewal. These include growth factors [e.g. FGF (fibroblast growth factor) and BMP (bone morphogenetic protein)], morphogens (e.g. Wnt), cell–cell contact/communication regulators (e.g. Notch), and mediators of the extracellular matrix (e.g. integrin- α 6) [1–3], all of which have been reported to control embryonic and adult stem cell homoeostasis. Other pathways have also been identified as modulators or co-modulators of stem cell function.

The p38 MAPK (mitogen-activated protein kinase) pathway is an important mediator of the cellular response to external signals, and in particular to stress. However, there have been many reports connecting this kinase pathway to the regulation of embryonic development and adult tissue turnover.

p38 MAPKs belong to the MAPK family. MAPKs can be classified into three groups: ERKs (extracellular-signal-regulated kinases); JNKs (c-Jun N-terminal kinases) and p38 MAPKs [4]. The present review will primarily focus on the role of p38 α in homoeostasis and disease.

p38 MAPKs have been considered as stress-activated protein kinases that respond to cellular stress and cytokines, with roles related to inflammation [5]. They can be divided into two subgroups, dependent on their expression pattern, substrate specificity and sensitivity to pharmacological inhibitors [6]. The

first group contains p38 α and p38 β , which are universally expressed, whereas the second group comprises p38 γ and p38 δ , which appear to have more tissue-specific expression patterns [7,8]. Strong activation of p38 MAPKs by cytokines and cellular stresses generally promotes the inhibition of cell growth and induces apoptosis [9,10,11]. The different p38 MAPK isoforms have been shown to have redundant, specific or even opposite functions, depending on the cell type involved and the nature of the stimulus [8,12]. The p38 α signalling pathway shows the typical kinase cascade of the MAPK family, which results in the regulation of a diverse range of cellular functions [13] (Figure 1).

 $p38\alpha$ MAPK is ubiquitously expressed and the most abundant member of the family. It is essential for embryonic development, while also regulating different cellular functions, including proliferation, differentiation, cell death, adhesion and migration, as well as the response to stress and metabolic pathways [14]. It does this through multiple mechanisms, including regulation of transcription, mRNA stability, chromatin remodelling and protein synthesis [14]. More recently, $p38\alpha$ has been found to play important roles in the maintenance of homoeostasis and related pathologies.

p38α IN DEVELOPMENT

p38 MAPKs are widely involved in development, regulating a plethora of processes, including growth, embryonic differentiation and tissue homoeostasis [15,16]. The role of p38 in development was first determined in *Drosophila* embryos, where disruption of p38 signalling by deletion of its upstream activator, MKK [MAPK kinase; lic (licorne)], caused mislocalization of Oskar mRNA and failure to position the embryos anterior–posterior and dorsal–ventral axes [17]. p38 α has also been found to play an essential role during development in

Abbreviations used: AP-1, activator protein 1; BMP, bone morphogenetic protein; C/EBP, CAAT/enhancer-binding protein; ESC, embryonic stem cell; IBD, inflammatory bowel disease; IFN, interferon; IL, interleukin; JNK, c-Jun N-terminal kinase; MAPK, mitogen-activated protein kinase; MEF, myocyte enhancer factor; MKK, MAPK kinase; MSC, mesenchymal stem cell; TGF, transforming growth factor; Th, T-cell helper; TNF, tumour necrosis factor.

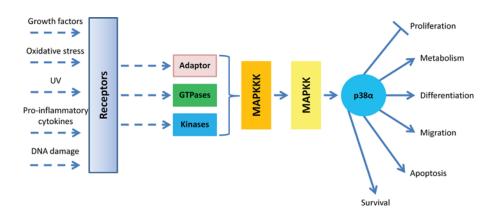


Figure 1 p38 α is part of the family of MAPKs

These kinase pathways are activated by external signals, and they form cascades of phosphorylation that lead to the activation of downstream targets and cellular responses. MAPKK, MKK; MAPKKK, MKK kinase

other animal models. In zebrafish, suppression of the p38 pathway affects the cleavage of the future dorsal side of the embryo and morphogenesis [18,19]. In *Xenopus*, the lack of p38 α affects early myogenic development, extending proliferation of the presomitic mesoderm and delaying somitogenesis [20]. In summary, p38 activity in non-mammalian organisms is essential for mesenchymal differentiation, which is required for the proper spatial organization of the animal during development.

Deletion of the mammalian p38 α isoform in murine embryos was shown to be lethal at E10.5 (embryonic day 10.5) [12,21,22]. Lethality was due to placental defects causing p38 $\alpha^{-/-}$ embryos to die from starvation and low oxygenation. Subsequently, the role of p38 α in trophoblast differentiation was demonstrated as the placental defect being overcome following fusion of $p38\alpha^{-/-}$ blastocysts with wild-type tetraploid cells (that only contribute to extraembryonic structures) [12]. Such rescue studies established an important role for the p38 α pathway during early placentation, but do not suggest that its activity is required for preimplantation development. Similarly, disruption of p38 signalling by compound deletion of the p38 upstream activators MKK3 and MKK6 in mice, results in embryonic death due to placental defects [23]. Furthermore, p38 α has been shown to be required for the development of the 8–16-cell stage of in vitro cultured embryos [24], regulating filamentous actin, as has also been demonstrated in zebrafish [19].

Elucidation of p38 MAPK functions from studies in knockout mice has provided valuable information on their relative importance during embryogenesis, but there is little knowledge about the developmental roles of p38 MAPKs at the cellular level. Embryonic lethality further limits in-depth analysis of the developmental role of p38 α at the cellular level in animal models, but generation of p38 $\alpha^{-/-}$ ESCs (embryonic stem cells) [25,26] has provided a valuable alternative system. It has been shown that cultured p38 $\alpha^{-/-}$ ESCs display several altered properties. These include augmented cell adhesion, which correlates with increased phosphorylation of focal adhesion kinase, and enhanced viability, owing to endogenous activation of Akt [27]. Induced differentiation of ESCs in vitro has shown that p38 α promotes mesodermal specification, whereas $p38\alpha^{-/-}$ ESCs tend to differentiate into neurons, reducing mesodermal commitment to a greater or lesser degree [28,29].

The apparent discrepancy between the dispensability of p38 α for embryo development in knockout animals [12,30,31] and the *in vitro* role for p38 α in ESC differentiation, can perhaps be explained by compensation via other p38 isoforms, namely p38 β , p38 γ or p38 δ . One might speculate that p38 β , which shares the

highest homology with p38 α [8], is the most likely candidate in this regard. It is also possible that differences in the *in vitro* set up, such as the use of serum containing undefined factors that would be able to compensate for the loss of function of p38 α , may result in the activation of other MAPK pathways [32].

In summary, these studies suggest that p38 α deletion may not significantly compromise the potential of ESC differentiation to certain cell types, but it may select for a commitment to specific cell lineages. p38 α is a likely activator of transcription factors not only involved in embryonic specification [e.g. Brachyury, MEF2C (myocyte enhancer factor 2C), PPAR (peroxisome-proliferator-activated receptor)] [29,33,34], but also in cross-talk with survival pathways (e.g. Akt) [27] and the regulation of proliferation mediators (e.g. cyclin D1) [35]. The balance between all of these effectors will induce different cellular activities depending on the state of development and cell type, producing apparently opposing outcomes.

More recently, the use of embryonic-specific conditional alleles has revealed specific functions for p38 α in later embryonic development, e.g. embryonic lung development, resulting in mice dying shortly after birth, most likely being due to disrupted differentiation of the bronchioalveolar epithelium [32]. Other reports have shown a role for p38 α in the branching of the developing lung, as blocking p38 signalling with the inhibitor SB203580, or following p38 α knockdown by shRNA (small hairpin RNA), suppressed budding morphogenesis of mouse embryonic lung explants [36]. The use of more specific conditional alleles will allow a better dissection of the functions and mediators of p38 α during different stages of embryonic development.

$p38\alpha$ In adult tissue homoeostasis

The use of conditional floxed alleles and specific small chemical inhibitors has allowed the study of the functions of $p38\alpha$ in adult tissues [14]. This approach has revealed important physiological functions for $p38\alpha$ in lung [37], liver [32], muscle [38], heart [39], haematopoiesis [40], skin [41] and pancreas [42]. In the present review we provide an overview of the recent advances in the study of the role of $p38\alpha$ in regulating adult tissue stem cell homoeostasis.

p38 α in haematopoiesis

The role of $p38\alpha$ as a central regulator of haematopoietic homoeostasis is well established. The regulation of proliferation, survival and differentiation of normal haematopoietic cells

by different cytokines and growth factors occurs via p38 α signalling [43–46]. In the haematopoietic system, p38 α exhibits contradictory functions in different lineages and maturation stages [47]. In thymic development, p38 α inhibits differentiation of immature thymocytes at specific stages [48]. In mature CD4+ cells, p38 α participates in Th1 (T-cell helper 1) inflammatory response, whereas it needs to be shut down in B-cell pro-activator Th2 cells [49,50].

A role for p38 α in mature granulocytes has also been reported. p38 α is essential for the survival of neutrophils during inflammation, and suppression of p38 α signalling is necessary to eliminate neutrophils in the termination of the inflammatory response [51,52]. The same activation/inactivation process is essential for the survival/apoptosis of eosinophils during the inflammatory response [53]. The function of p38 α activity in myeloid cells is paradoxical. Whereas in CD34⁺ progenitor cells, elevated p38α activity prevents haematopoiesis [54], in other myeloid cells, p38 α mediates activin A-mediated differentiation. These opposite effects may be due to the role of p38 α as a regulator of transcription factors involved in differentiation and its role in cytokine expression in response to stress [55]. The roles of p38 α in haematopoiesis are mediated by its activation of different pools of cell-dependent cytokines [49] or transcription factors [e.g. C/EBP (CAAT/enhancer-binding protein) and GATA-1] [56,57], or mediating differentiation signals [58,59]. The level of p38 α activation together with cross-talking to other pathways results in diverse outcomes, such as apoptosis, survival, differentiation or progenitor proliferation [58]. A better understanding of the specific roles of p38 α in every stage of haematopoietic development will help in tackling blood-related diseases.

p38 α in muscle regeneration

The p38 α MAPK pathway is an important regulator of skeletal muscle differentiation (myogenesis) [60,61]. The regulation of myogenesis is essential for normal development, as well as being important in pathological processes (e.g. muscular dystrophies and inflammatory myopathies) in which marked muscle loss and regeneration occurs. The regenerative capacity of adult skeletal muscle has been demonstrated upon acute muscle damage, resulting in the post-trauma generation of myotubes after a few days [62,63]. Early research suggested that budding of myotubes from injured myofibres was the source of new myofibres [63]. Satellite cells have been suggested as being the source for 'dormant myoblasts', responding to muscle damage by reinitiating a process similar to skeletal myogenesis [64]. Satellite cells are activated from quiescence by the p38 MAPK pathway [65].

 $p38\alpha$ participates in various stages of adult myogenic differentiation [66]. At early stages, $p38\alpha$ promotes the active heterodimer transcription complex, MyoD/E47 and phosphorylates MEF2, inducing the expression of muscle-specific genes and thereby activating the differentiation program [67,68]. However, at later stages of myogenic differentiation, $p38\alpha$ plays a suppressive role. Phosphorylation of the myogenic factor MRF4 (muscle-regulatory factor 4) reduces its transcriptional activity, affecting essential genes involved in terminal differentiation [69,70]. Nevertheless, the suppressive role of $p38\alpha$ in late myogenesis has only been demonstrated *in vitro* and it requires validation *in vivo*.

The regulation of myoblast proliferation by $p38\alpha$ represents a novel role for the p38 pathway in skeletal myogenesis [38]. $p38\alpha^{-/-}$ myoblasts are characterized by their increased proliferation, a delay in cell-cycle exit, and impaired myoblast differentiation and fusion. $p38\alpha$ is the central p38 MAPK

responsible for both *in vitro* and *in vivo* regulation of myogenesis [71].

A key role for p38 α in controlling myoblast proliferation is the antagonism of the JNK/c-Jun pathway, probably via MKP-1 (MAPK phosphatase-1) [72]. The cross-talk between the p38 MAPK and JNK signalling pathways, by still undefined mechanisms, has been previously described in different cell types [73–76]. In the context of skeletal myogenesis, two studies have suggested opposite roles for JNK activity in muscle differentiation [77,78]. Importantly, JNK activation has been shown to mediate the increased proliferation potential of p38 α -deficient myoblasts, with inhibition of JNK reverting this phenotype. Moreover, enhanced activation of JNK in p38 α -deficient myoblasts results in increased levels of its substrate phospho-c-Jun and subsequent induction of c-Jun/AP-1 (activator protein 1)-mediated c-Jun gene transcription. This leads to increased recruitment of c-Jun to the cyclin D1 loci in differentiating myoblasts in vivo, presumably via the AP-1 sites on the cyclin D1 promoter [79]. p38α controls myoblast proliferation by antagonizing the proproliferative activity of JNK [80].

 $p38\alpha$ appears to play contradictory roles in muscle differentiation (Figure 2). The availability of specific transcription factors at particular stages of myogenesis, and the effect (activating or inactivating) that $p38\alpha$ -dependent phosphorylation induces, together with the cross-talk with other MAPK pathways, may be responsible for promoting or suppressing differentiation at early or late stages of myogenesis.

$p38\alpha$ in neurogenesis

Most of the known roles of $p38\alpha$ in neurogenesis are related to embryonic development. The roles of p38 in early ESC commitment are due to various insults triggering differentiation that may involve p38 targets, such as repression of Bcl2 expression, leading to neural differentiation [81], or induction of BMP2 mRNA, to mesodermal [82] differentiation.

In later stages of embryonic development, p38 activity prevents neurogenesis (probably due to p38 β activity) in embryonic cortical [83] and oxygen/glucose-deprived hippocampal neurons [84]. However, proliferation of adult hippocampal neural progenitor cells is dependent on adiponectin activation of a p38/GSK3 (glycogen synthase kinase 3)/ β -catenin cascade [85]. Furthermore, p38 signalling promotes adult neural differentiation by activating neural transcription factors such as neurogenin 1 [86] and oligodendrocyte progenitor cell progression through Sox10 [SRY (sex-determining region Y)-box 10] activation [87]. The p38 α /MEF2C pathway is a survival signal during neural differentiation [88]. In addition, p38 activation by TGF (transforming growth factor)- β promotes differentiation of retinal ganglion cells and neurite outgrowth [89].

Neurogenesis is a clear example of the opposing functions that $p38\alpha$ can show in the same organ. The context, as a combination of the differentiation stage of the cells and the external signals from the environment, will influence the positive or negative outcome of p38 signalling in neural differentiation.

$p38\alpha$ in cardiac homoeostasis

The heart undergoes the least amount of tissue turnover, and regeneration has only been shown in a few species (e.g. newts and zebrafish). Following myocardial infarction, it has been suggested that more than one billion cardiomyocytes per patient would be required for tissue replacement therapy [90]. The zebrafish model has provided a good system to study cardiac self-repair [91,92].

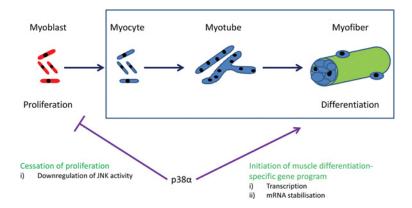


Figure 2 Muscle differentiation model as a paradigm of p38 α -dependent tissue stem cell regulation

 $p38\alpha$ promotes differentiation and suppresses self-renewal of myoblast progenitors.

The hypothesis of a terminal round of cell division, resulting in the majority of cardiac myocytes exiting the cell cycle shortly after birth [93,94], led to the belief that the mammalian heart was terminally differentiated and lacked the capacity for myocardial self-renewal following injury. However, the identification of cardiac progenitor cells has led to the belief that the heart is also regulated by an adult stem cell compartment [95–101].

 $p38\alpha$ has been shown to be a key regulator of mammalian cardiomyogenesis. This includes roles in cardiomyocyte differentiation, division, apoptosis and hypertrophy [102]. Furthermore, p38 MAPK has been shown to control myoblast differentiation at multiple levels, including regulation of transcription factor activity, chromatin remodelling and stability of mRNAs encoding muscle differentiation regulators. p38 α regulates cardiac differentiation transcription factors, such as GATA4 [103], MEF2C [104] and SRF (serum-response factor) [105]. Furthermore, it has also been suggested that C/EBP β and TEF-1 (transcriptional enhancer factor 1) mediate p38 MAPK function in cardiomyocytes following myocardial injury [106], thereby playing a central role in the repair response.

 $p38\alpha$ activity in post-injury hearts plays an important role in cellular and myocardial remodelling, affecting both the contractility of myocytes and the extracellular matrix. Dissecting the underlying mechanisms involved in the myocyte cell, autonomous effects as well as the cross-talk interaction between myocytes, fibroblasts and inflammatory cells, should provide a very promising area for future investigation.

p38 α in other tissues

p38 also plays roles in other mesenchymal tissues. It has been extensively reported that p38 plays a role in bone regeneration and repair [107]. As in other cellular contexts, the observed roles of p38 α in bone differentiation may be contradictory depending on the activation of p38 α as an inflammatory mediator or as a regulator of differentiation factors. p38, upon activation by chondrogenic cytokines in MSCs (mesenchymal stem cells), is an essential mediator of bone formation [107–109]. Osteoblast differentiation is regulated by p38 MAPK activation of RUNX1 (Runt-related transcription factor 1) [110]. However, p38 α is directly involved in osteoclast differentiation and bone-resorbing activity induced by inflammation [111]. In adipose tissue, there are also opposing reports identifying p38 α as an inhibitor [34] or inducer [112] of adipocyte differentiation. In general, early activity of p38 in MSCs promotes bone differentiation [113], but

a later activation of the p38 pathway in MSCs leads to their specification into white or brown adipocytes [112,114].

The p38 α pathway is also essential for mature differentiation of epithelial tissues. Differentiation of human pancreatic islets regulated by TGF- β or activin A is dependent on p38 activation of the transcription factor PAX6 (paired box 6) [115,116]. The MKK3/p38 cascade regulates the expression of specific pancreatic factors, for example neurogenin-3, which allows pancreatic endocrine cell differentiation [117].

Intestinal homoeostasis can be disrupted by specific deletion of p38 α in the colonic epithelium [55]. This intestinal function of p38 α maybe mediated through the induction of intestinal genes such as Schlafen-3 [118].

There is also increasing evidence for a pivotal role of $p38\alpha$ at various levels of lung differentiation. $p38\alpha$ is essential in late lung development differentiation, and lack of p38 causes neonatal lethality [32,119]. This role is conserved in the adult lung, and $p38\alpha$ is necessary to maintain adult lung homoeostasis [37]. Furthermore, although p38 can be involved in lung regeneration induced by acute inflammation [120], it is also a mediator of TGF- β -dependent epithelial to mesenchymal transition in bleomycin-induced lung fibrosis [121].

In mammary development, p38 α plays a negative role in differentiation by reducing the proliferation needed for ductal expansion and branching morphogenesis [122].

Regulation of caspase 14 in skin by p38 is necessary for normal differentiation of epidermal keratinocytes [123], and shape-induced terminal differentiation of skin stem cells needs p38 α activity [124].

In general, the role of p38 α in adult epithelial tissues is determined by the prevalence of its inflammatory functions (usually directing to tissue injury) or the activation of differentiation factors allowing tissue homoeostasis.

$p38\alpha$ DISRUPTION IN DISEASE

Cellular behaviour in response to extracellular stimuli is mediated through intracellular signalling pathways, such as the p38 MAPK pathways, and abnormal phosphorylation events can be a cause of, or contribute towards, disease progression in a variety of disorders (Figure 3). The best-known and most widely reported role of p38 α in disease is related to its function in cytokine signalling and promotion of pathological inflammation [125]. Several disease models, including rheumatoid arthritis, psoriasis, Alzheimer's disease [126], IBD (inflammatory bowel disease) [127,128],

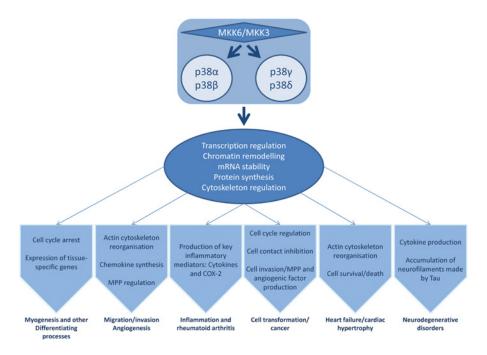


Figure 3 Diversity of cellular processes and physiological functions regulated by the p38 MAPK family

The pathological consequences of the abnormal activation of the pathway are shown at the bottom of the Figure, COX-2, cyclo-oxygenase 2; MPP, Mg²⁺-dependent protein phosphatase.

Crohn's disease [129], tumorigenesis [14], cardiovascular disease and stroke [130] are all postulated to be mediated, at least in part, by the p38 α pathway. Furthermore, previous studies support a role for p38 MAPK in the development, maintenance and/or exacerbation of a number of pulmonary diseases, such as asthma, cystic fibrosis, idiopathic pulmonary fibrosis and COPD (chronic obstructive pulmonary disease) [131]. However, evidence highlights the importance of the differentiation roles of p38 α in relation to pathological processes [132].

Many of the inflammation-dependent diseases associated with p38 α activity are linked to cytokine-induced differentiation [133]. Activation of p38 MAPK signalling mediates direct or indirect inflammatory cytokine expression, such as IL (interleukin)- 1β , IL-6 and TNF (tumour necrosis factor)- α . These cytokines synergistically stimulate the production of other inflammatory cytokines, MMPs (matrix metalloproteinases) and prostanoids [134,135]. p38 α has also been involved in the regulation of IL-3, IL-8, MIP1 α (macrophage inhibitory protein 1α), GM-CSF (granulocyte/macrophage colony-stimulating factor), VEGF (vascular endothelial growth factor), urokinase-type plasminogen activator and inducible nitric oxide synthase [13,21]. The duration of phosphorylation is crucial in regulating cell fate. Sustained p38 α phosphorylation is frequently associated with induction of cellular apoptosis [136,137]; in contrast, transient phosphorylation can be associated with growth-factor-induced survival [138]. Thus the opposite cellular outcome may influence the advance or regression in pathological states.

TNF production in the bone is induced by p38 in rheumatoid arthritis and responsible for bone destruction and inhibition of chondrocyte differentiation, promoting fibrosis in the damaged tissue [139,140]. Increased levels of p38 α phosphorylation are seen in the epidermal cells of psoriatic lesions, playing an essential role in promoting the characteristic flaky skin [123]. In asthma, p38 and its downstream target MAPKAPK2 (MAPK-activated protein kinase) are involved in type 2 Th2 cell final activation and differentiation, and production of MCP (monocyte chemoattractant protein)-1 by lung epithelial cells [141,142].

Activation of p38 MAPK induces pro-inflammatory cytokines in IBD, such as IL-1 β and TNF- α , both in production and secretion. This regulation takes place in non-immune cells, such as HIMECs (human intestinal microvascular endothelial cells), intestinal epithelium, fibroblasts and myofibroblasts, which participate in IBD and are subject to the direct or indirect effect of p38 MAPK [143,144], and in immune cells such as monocytes and macrophages [145]. Chronic p38 α -mediated inflammatory events disrupt homoeostasis and prevent epithelial differentiation, consequently promoting fibroblast maturation and proliferation.

The role of p38 α in the neural system has been best demonstrated in relation to neurodegenerative diseases [132]. Disrupted p38 activation has been shown in animal models of neurodegeneration [126] and deposition of tau protein in related pathologies (e.g. Alzheimer's disease) [146]. Although some of the functions are due to the control of inflammatory cytokines released by p38 [147], it has been found that p38 α can phosphorylate tau, thereby reducing its ability to promote microtubule assembly [148,149]. In addition, p38 α expression in spinal cord neurons seems to be related to neuropathic pain [150,151]. It has also been shown that p38 mediates the survival of cerebellar granule neurons [152].

The p38 MAPK pathway can have both protective and detrimental effects in cardiovascular diseases [132]. It plays an important role in cardiovascular remodelling after injury [153]. After myocardial infarction, p38 activation is a negative regulator of cardiomyocyte proliferation [154]. This potentiates tissue apoptosis [155] and promotes fibrosis [156,157]. Conversely, p38 α induces proliferation of vascular smooth muscle cells and vascular regeneration after carotid injury [153].

The complexity of the functions of $p38\alpha$ in the maintenance of haematopoiesis influences its role in haematopoietic diseases. On one hand, $p38\alpha$ is responsible for enhanced stem cell apoptosis, a characteristic of low-grade myelodysplastic syndromes [54,158]. On the other hand, an imbalance towards proliferation may lead to the development of myeloproliferative syndromes, such as leukaemia, lymphomas and myelomas [159–161]. p38 MAPK

is selectively activated by IFN α (interferon α) and mediates the growth-suppressive effects of IFN α in CML (chronic myeloid leukaemia) cells [162].

Finally, the variety of cellular processes involving p38 include many that oppose the oncogenic transformation of solid tissues. As a result, p38 α has been considered a tumour suppressor [163,164]. Although most of the suppressor activity is apparently due to promotion of growth arrest and induction of apoptosis [165,166], p38 also contributes to the loss of a malignant phenotype by inducing terminal differentiation of solid epithelial cancer cells [37,167,168].

The paradoxical and contradictory effects of $p38\alpha$ in disease are again closely related to its functions as an inflammatory or differentiation mediator.

CONCLUDING REMARKS

A broad range of intracellular mediators and extracellular insults are involved in $p38\alpha$ activation and function in the cell. They are responsible for the contradictory roles of this pathway not only in different tissues, but also within the same organs and cell types. These opposite roles seem to be related to duration and level of kinase activity. In general, long-term and high levels of $p38\alpha$ activity are involved in the inflammatory response, which usually leads to promotion and progression of disease. High, but transient activation, is linked to apoptosis and may suppress disease (e.g. cancer) or promote pathological processes (e.g. cardiomyopathy). However, constitutive low-level activity promotes differentiation and negatively regulates proliferation. It is this marginal but constant activity that has been reported as being essential for correct stem cell regulation.

Many p38 inhibitors have been developed to tackle inflammatory diseases [6]. Inhibition of p38 prevents the response to inflammatory cytokines and cytokine production at the same time. However, many trials have been stopped owing to toxicities in several tissues. This may be due to the variety of p38 α functions and the non-specific cellular inhibition by those drugs [169]. Inhibition of constitutive p38 activity may also interfere with proper cellular turnover and organ physiology. Investigation of the cellular and functional roles of p38 α in specific physiological and pathological processes in every organ will allow a better understanding of the responses to drugs targeting this kinase pathway. Cellular and molecular specific drugs directed against mediators of the p38 α signalling will improve future use of chemical inhibitors of this pathway in disease therapy.

ACKNOWLEDGEMENTS

We thank E. Hoste and D. Winder for critically reading the paper prior to submission.

FUNDING

This group is funded by the Medical Research Council (MRC) [grant numbers RG51968 and RG57589] and Cancer Research UK [grant number RG52191].

REFERENCES

- 1 Czyz, J. and Wobus, A. (2001) Embryonic stem cell differentiation: the role of extracellular factors. Differentiation 68, 167–174
- 2 Iglesias-Bartolome, R. and Gutkind, J. S. (2011) Signaling circuitries controlling stem cell fate: to be or not to be. Curr. Opin. Cell Biol. 23, 716–723
- 3 Watt, F. M. (2002) Role of integrins in regulating epidermal adhesion, growth and differentiation. EMBO J. 21, 3919–3926

- 4 Raman, M., Chen, W. and Cobb, M. H. (2007) Differential regulation and properties of MAPKs. Oncogene 26, 3100–3112
- 5 Nebreda, A. R. and Porras, A. (2000) p38 MAP kinases: beyond the stress response. Trends Biochem. Sci. 25, 257–260
- 6 Cohen, P. (2009) Targeting protein kinases for the development of anti-inflammatory drugs. Curr. Opin. Cell Biol. 21, 317–324
- 7 Jiang, Y., Li, Z., Schwarz, E. M., Lin, A., Guan, K., Ulevitch, R. J. and Han, J. (1997) Structure-function studies of p38 mitogen-activated protein kinase. Loop 12 influences substrate specificity and autophosphorylation, but not upstream kinase selection. J. Biol. Chem. 272, 11096–11102
- 8 Ono, K. and Han, J. (2000) The p38 signal transduction pathway: activation and function. Cell. Signalling 12, 1–13
- 9 Kyriakis, J. M. and Avruch, J. (1996) Protein kinase cascades activated by stress and inflammatory cytokines. Bioessays 18, 567–577
- 10 Kyriakis, J. M. and Avruch, J. (2001) Mammalian mitogen-activated protein kinase signal transduction pathways activated by stress and inflammation. Physiol. Rev. 81, 807–869
- 11 Guo, J. H., Wang, H. Y. and Malbon, C. C. (1998) Conditional, tissue-specific expression of Q205L Gαi2 *in vivo* mimics insulin activation of c-Jun N-terminal kinase and p38 kinase. J. Biol. Chem. **273**, 16487–16493
- 12 Adams, R. H., Porras, A., Alonso, G., Jones, M., Vintersten, K., Panelli, S., Valladares, A., Perez, L., Klein, R. and Nebreda, A. R. (2000) Essential role of p38α MAP kinase in placental but not embryonic cardiovascular development. Mol. Cell 6, 109–116
- 13 Cobb, M. H. and Goldsmith, E. J. (1995) How MAP kinases are regulated. J. Biol. Chem. 270, 14843—14846
- 14 Cuadrado, A. and Nebreda, A. R. (2010) Mechanisms and functions of p38 MAPK signalling. Biochem. J. 429, 403–417
- 15 Rose, B. A., Force, T. and Wang, Y. (2010) Mitogen-activated protein kinase signaling in the heart: angels versus demons in a heart-breaking tale. Physiol. Rev. 90, 1507–1546
- 16 Roux, P. P. and Blenis, J. (2004) ERK and p38 MAPK-activated protein kinases: a family of protein kinases with diverse biological functions. Microbiol. Mol. Biol. Rev. 68, 320–344
- 17 Suzanne, M., Irie, K., Glise, B., Agnes, F., Mori, E., Matsumoto, K. and Noselli, S. (1999) The *Drosophila* p38 MAPK pathway is required during oogenesis for egg asymmetric development. Genes Dev. 13, 1464–1474
- 18 Fujii, R., Yamashita, S., Hibi, M. and Hirano, T. (2000) Asymmetric p38 activation in zebrafish: its possible role in symmetric and synchronous cleavage. J. Cell. Biol. 150, 1335–1348
- Holloway, B. A., Gomez de la Torre Canny, S., Ye, Y., Slusarski, D. C., Freisinger, C. M., Dosch, R., Chou, M. M., Wagner, D. S. and Mullins, M. C. (2009) A novel role for MAPKAPK2 in morphogenesis during zebrafish development. PLoS Genet. 5, e1000413
- 20 Keren, A., Bengal, E. and Frank, D. (2005) p38 MAP kinase regulates the expression of XMyf5 and affects distinct myogenic programs during *Xenopus* development. Dev. Biol. 288, 73–86
- 21 Allen, M., Svensson, L., Roach, M., Hambor, J., McNeish, J. and Gabel, C. A. (2000) Deficiency of the stress kinase p38α results in embryonic lethality: characterization of the kinase dependence of stress responses of enzyme-deficient embryonic stem cells. J. Exp. Med. 191, 859–870
- 22 Mudgett, J. S., Ding, J., Guh-Siesel, L., Chartrain, N. A., Yang, L., Gopal, S. and Shen, M. M. (2000) Essential role for p38α mitogen-activated protein kinase in placental angiogenesis. Proc. Natl. Acad. Sci. U.S.A. 97, 10454–10459
- 23 Brancho, D., Tanaka, N., Jaeschke, A., Ventura, J. J., Kelkar, N., Tanaka, Y., Kyuuma, M., Takeshita, T., Flavell, R. A. and Davis, R. J. (2003) Mechanism of p38 MAP kinase activation in vivo. Genes Dev. 17. 1969–1978
- 24 Natale, D. R., Paliga, A. J., Beier, F., D'Souza, S. J. and Watson, A. J. (2004) p38 MAPK signaling during murine preimplantation development. Dev. Biol. 268, 76–88
- 25 Han, J., Richter, B., Li, Z., Kravchenko, V. and Ulevitch, R. J. (1995) Molecular cloning of human p38 MAP kinase. Biochim. Biophys. Acta 1265, 224–227
- 26 Kim, S. J., Ko, C. B., Park, C., Kim, B. R., Sung, T. H., Koh, D. H., Kim, N. S., Oh, K. J., Chung, S. Y. and Park, R. (2005) p38 MAP kinase regulates benzo(a)pyrene-induced apoptosis through the regulation of p53 activation. Arch. Biochem. Biophys. 444, 121–120
- 27 Guo, Y. L. and Yang, B. (2006) Altered cell adhesion and cell viability in a p38 α mitogen-activated protein kinase-deficient mouse embryonic stem cell line. Stem Cells Dev. **15**, 655–664
- 28 Guo, Y. L., Ye, J. and Huang, F. (2007) p38 α MAP kinase-deficient mouse embryonic stem cells can differentiate to endothelial cells, smooth muscle cells and neurons. Dev. Dyn. **236**, 3383–3392
- 29 Barruet, E., Hadadeh, O., Peiretti, F., Renault, V. M., Hadjal, Y., Bernot, D., Tournaire, R., Negre, D., Juhan-Vague, I., Alessi, M. C. et al. (2011) p38 mitogen activated protein kinase controls two successive-steps during the early mesodermal commitment of embryonic stem cells. Stem Cells Dev. 20, 1233–1246

- 30 Beardmore, V. A., Hinton, H. J., Eftychi, C., Apostolaki, M., Armaka, M., Darragh, J., McIlrath, J., Carr, J. M., Armit, L. J., Clacher, C. et al. (2005) Generation and characterization of p38β (MAPK11) gene-targeted mice. Mol. Cell Biol. 25, 10454–10464
- 31 Sabio, G., Arthur, J. S., Kuma, Y., Peggie, M., Carr, J., Murray-Tait, V., Centeno, F., Goedert, M., Morrice, N. A. and Cuenda, A. (2005) p38y regulates the localisation of SAP97 in the cytoskeleton by modulating its interaction with GKAP. EMBO J. 24, 1134–1145
- 32 Hui, L., Bakiri, L., Mairhorfer, A., Schweifer, N., Haslinger, C., Kenner, L., Komnenovic, V., Scheuch, H., Beug, H. and Wagner, E. F. (2007) p38α suppresses normal and cancer cell proliferation by antagonizing the JNK-c-Jun pathway. Nat. Genet. 39, 741–749
- 33 Aouadi, M., Bost, F., Caron, L., Laurent, K., Le Marchand Brustel, Y. and Binetruy, B. (2006) p38 mitogen-activated protein kinase activity commits embryonic stem cells to either neurogenesis or cardiomyogenesis. Stem Cells 24, 1399–1406
- 34 Aouadi, M., Laurent, K., Prot, M., Le Marchand-Brustel, Y., Binetruy, B. and Bost, F. (2006) Inhibition of p38MAPK increases adipogenesis from embryonic to adult stages. Diabetes 55, 281–289
- 35 Awad, M. M., Enslen, H., Boylan, J. M., Davis, R. J. and Gruppuso, P. A. (2000) Growth regulation via p38 mitogen-activated protein kinase in developing liver. J. Biol. Chem. 275, 38716–38721
- 36 Liu, Y., Martinez, L., Ebine, K. and Abe, M. K. (2008) Role for mitogen-activated protein kinase p38α in lung epithelial branching morphogenesis. Dev. Biol. 314, 224–235
- 37 Ventura, J. J., Tenbaum, S., Perdiguero, E., Huth, M., Guerra, C., Barbacid, M., Pasparakis, M. and Nebreda, A. R. (2007) p38α MAP kinase is essential in lung stem and progenitor cell proliferation and differentiation. Nat. Genet. 39, 750–758
- 38 Perdiguero, E., Ruiz-Bonilla, V., Gresh, L., Hui, L., Ballestar, E., Sousa-Victor, P., Baeza-Raja, B., Jardi, M., Bosch-Comas, A., Esteller, M. et al. (2007) Genetic analysis of p38 MAP kinases in myogenesis: fundamental role of p38α in abrogating myoblast proliferation. EMBO J. 26, 1245–1256
- 39 Engel, F. B., Schebesta, M., Duong, M. T., Lu, G., Ren, S., Madwed, J. B., Jiang, H., Wang, Y. and Keating, M. T. (2005) p38 MAP kinase inhibition enables proliferation of adult mammalian cardiomyocytes. Genes Dev. 19, 1175–1187
- 40 Kang, Y. J., Chen, J., Otsuka, M., Mols, J., Ren, S., Wang, Y. and Han, J. (2008) Macrophage deletion of p38α partially impairs lipopolysaccharide-induced cellular activation. J. Immunol. 180, 5075–5082
- 41 Kim, C., Sano, Y., Todorova, K., Carlson, B. A., Arpa, L., Celada, A., Lawrence, T., Otsu, K., Brissette, J. L., Arthur, J. S. et al. (2008) The kinase p38α serves cell type-specific inflammatory functions in skin injury and coordinates pro- and anti-inflammatory gene expression. Nat. Immunol. 9, 1019–1027
- 42 Wong, E. S., Le Guezennec, X., Demidov, O. N., Marshall, N. T., Wang, S. T., Krishnamurthy, J., Sharpless, N. E., Dunn, N. R. and Bulavin, D. V. (2009) p38MAPK controls expression of multiple cell cycle inhibitors and islet proliferation with advancing age. Dev. Cell 17, 142–149
- 43 Nagata, Y., Moriguchi, T., Nishida, E. and Todokoro, K. (1997) Activation of p38 MAP kinase pathway by erythropoietin and interleukin-3. Blood 90, 929–934
- 44 Verma, A., Deb, D. K., Sassano, A., Uddin, S., Varga, J., Wickrema, A. and Platanias, L. C. (2002) Activation of the p38 mitogen-activated protein kinase mediates the suppressive effects of type I interferons and transforming growth factor-β on normal hematopoiesis. J. Biol. Chem. 277. 7726–7735
- 45 Katsoulidis, E., Li, Y., Mears, H. and Platanias, L. C. (2005) The p38 mitogen-activated protein kinase pathway in interferon signal transduction. J. Interferon Cytokine Res. 25, 749–756
- 46 Jacobs-Helber, S. M., Ryan, J. J. and Sawyer, S. T. (2000) JNK and p38 are activated by erythropoietin (EPO) but are not induced in apoptosis following EPO withdrawal in EPO-dependent HCD57 cells. Blood 96, 933–940
- 47 Rincon, M. and Pedraza-Alva, G. (2003) JNK and p38 MAP kinases in CD4 + and CD8 + T cells. Immunol. Rev. 192, 131–142
- 48 Diehl, N. L., Enslen, H., Fortner, K. A., Merritt, C., Stetson, N., Charland, C., Flavell, R. A., Davis, R. J. and Rincon, M. (2000) Activation of the p38 mitogen-activated protein kinase pathway arrests cell cycle progression and differentiation of immature thymocytes in vivo. J. Exp. Med. 191, 321–334
- 49 Rincon, M., Enslen, H., Raingeaud, J., Recht, M., Zapton, T., Su, M. S., Penix, L. A., Davis, R. J. and Flavell, R. A. (1998) Interferon-γ expression by Th1 effector T cells mediated by the p38 MAP kinase signaling pathway. EMBO J. 17, 2817–2829
- 50 Merritt, C., Enslen, H., Diehl, N., Conze, D., Davis, R. J. and Rincon, M. (2000) Activation of p38 mitogen-activated protein kinase *in vivo* selectively induces apoptosis of CD8+ but not CD4+ T cells. Mol. Cell Biol. 20, 936–946
- 51 Alvarado-Kristensson, M., Melander, F., Leandersson, K., Ronnstrand, L., Wernstedt, C. and Andersson, T. (2004) p38-MAPK signals survival by phosphorylation of caspase-8 and caspase-3 in human neutrophils. J. Exp. Med. 199, 449–458

- 52 Alvarado-Kristensson, M., Porn-Ares, M. I., Grethe, S., Smith, D., Zheng, L. and Andersson, T. (2002) p38 mitogen-activated protein kinase and phosphatidylinositol 3-kinase activities have opposite effects on human neutrophil apoptosis. FASEB J. 16, 129–131
- 53 Kankaanranta, H., De Souza, P. M., Barnes, P. J., Salmon, M., Giembycz, M. A. and Lindsay, M. A. (1999) SB 203580, an inhibitor of p38 mitogen-activated protein kinase, enhances constitutive apoptosis of cytokine-deprived human eosinophils. J. Pharmacol Exp. Ther. 290, 621–628
- 54 Navas, T. A., Mohindru, M., Estes, M., Ma, J. Y., Sokol, L., Pahanish, P., Parmar, S., Haghnazari, E., Zhou, L., Collins, R. et al. (2006) Inhibition of overactivated p38 MAPK can restore hematopoiesis in myelodysplastic syndrome progenitors. Blood 108, 4170–4177
- 55 Otsuka, M., Kang, Y. J., Ren, J., Jiang, H., Wang, Y., Omata, M. and Han, J. (2010) Distinct effects of p38α deletion in myeloid lineage and gut epithelia in mouse models of inflammatory bowel disease. Gastroenterology 138, 1255–1265
- 56 Buck, I., Morceau, F., Cristofanon, S., Heintz, C., Chateauvieux, S., Reuter, S., Dicato, M. and Diederich, M. (2008) Tumor necrosis factor α inhibits erythroid differentiation in human erythropoietin-dependent cells involving p38 MAPK pathway, GATA-1 and F0G-1 downregulation and GATA-2 upregulation. Biochem. Pharmacol. 76, 1229–1239
- 57 Geest, C. R., Buitenhuis, M., Laarhoven, A. G., Bierings, M. B., Bruin, M. C., Vellenga, E. and Coffer, P. J. (2009) p38 MAP kinase inhibits neutrophil development through phosphorylation of C/EBP α on serine 21. Stem Cells **27**, 2271–2282
- 58 Kale, V. P. (2004) Differential activation of MAPK signaling pathways by TGF-β1 forms the molecular mechanism behind its dose-dependent bidirectional effects on hematopoiesis. Stem Cells Dev. 13, 27–38
- 59 Vijayaraj, P., Kroeger, C., Reuter, U., Hartmann, D. and Magin, T. M. (2010) Keratins regulate yolk sac hematopoiesis and vasculogenesis through reduced BMP-4 signaling. Eur. J. Cell Biol. 89, 299–306
- 60 Keren, A., Tamir, Y. and Bengal, E. (2006) The p38 MAPK signaling pathway: a major regulator of skeletal muscle development. Mol. Cell. Endocrinol. 252, 224–230
- 61 Lluis, F., Perdiguero, E., Nebreda, A. R. and Munoz-Canoves, P. (2006) Regulation of skeletal muscle gene expression by p38 MAP kinases. Trends Cell Biol. 16, 36–44
- 62 Adams, R. D. and Walton, J. N. (1956) The response of the normal, the denervated and the dystrophic muscle-cell to injury. J. Pathol. Bacteriol. 72, 273–298
- 63 Legros, J. (1946) [Not Available]. Acta Paediatr. Belg. 1, 80–82
- 64 Mauro, A. (1961) Satellite cell of skeletal muscle fibers. J. Biophys. Biochem. Cytol. 9, 493–495
- 65 Jones, N. C., Tyner, K. J., Nibarger, L., Stanley, H. M., Cornelison, D. D., Fedorov, Y. V. and Olwin, B. B. (2005) The p38 α/β MAPK functions as a molecular switch to activate the quiescent satellite cell. J. Cell Biol. **169**, 105–116
- 66 Wu, Z., Woodring, P. J., Bhakta, K. S., Tamura, K., Wen, F., Feramisco, J. R., Karin, M., Wang, J. Y. and Puri, P. L. (2000) p38 and extracellular signal-regulated kinases regulate the myogenic program at multiple steps. Mol. Cell Biol. 20, 3951–3964
- 67 Lluis, F., Ballestar, E., Suelves, M., Esteller, M. and Munoz-Canoves, P. (2005) E47 phosphorylation by p38 MAPK promotes MyoD/E47 association and muscle-specific gene transcription. EMBO J. 24, 974–984
- 68 Zetser, A., Gredinger, E. and Bengal, E. (1999) p38 mitogen-activated protein kinase pathway promotes skeletal muscle differentiation. Participation of the Mef2c transcription factor. J. Biol. Chem. 274, 5193–5200
- 69 Suelves, M., Lluis, F., Ruiz, V., Nebreda, A. R. and Munoz-Canoves, P. (2004) Phosphorylation of MRF4 transactivation domain by p38 mediates repression of specific myogenic genes. EMBO J. 23, 365–375
- 70 Weston, A. D., Sampaio, A. V., Ridgeway, A. G. and Underhill, T. M. (2003) Inhibition of p38 MAPK signaling promotes late stages of myogenesis. J. Cell Sci. 116, 2885–2893
- 71 Ruiz-Bonilla, V., Perdiguero, E., Gresh, L., Serrano, A. L., Zamora, M., Sousa-Victor, P., Jardi, M., Wagner, E. F. and Munoz-Canoves, P. (2008) Efficient adult skeletal muscle regeneration in mice deficient in p38 β , p38 γ and p38 δ MAP kinases. Cell Cycle **7**, 2208–2214
- 72 Perdiguero, E., Sousa-Victor, P., Ruiz-Bonilla, V., Jardi, M., Caelles, C., Serrano, A. L. and Munoz-Canoves, P. (2011) p38/MKP-1-regulated AKT coordinates macrophage transitions and resolution of inflammation during tissue repair. J. Cell Biol. 195, 307–322
- 73 Chen, C. Y., Gherzi, R., Andersen, J. S., Gaietta, G., Jurchott, K., Royer, H. D., Mann, M. and Karin, M. (2000) Nucleolin and YB-1 are required for JNK-mediated interleukin-2 mRNA stabilization during T-cell activation. Genes Dev. 14, 1236–1248
- 74 Nemoto, S., Sheng, Z. and Lin, A. (1998) Opposing effects of Jun kinase and p38 mitogen-activated protein kinases on cardiomyocyte hypertrophy. Mol. Cell Biol. 18, 3518–3526
- 75 Porras, A., Zuluaga, S., Black, E., Valladares, A., Alvarez, A. M., Ambrosino, C., Benito, M. and Nebreda, A. R. (2004) p38α mitogen-activated protein kinase sensitizes cells to apoptosis induced by different stimuli. Mol. Biol. Cell 15, 922–933

- 76 Zechner, D., Craig, R., Hanford, D. S., McDonough, P. M., Sabbadini, R. A. and Glembotski, C. C. (1998) MKK6 activates myocardial cell NF-κB and inhibits apoptosis in a p38 mitogen-activated protein kinase-dependent manner. J. Biol. Chem. 273, 8232–8239
- 77 Khurana, A. and Dey, C. S. (2004) Involvement of c-Jun N-terminal kinase activities in skeletal muscle differentiation. J. Muscle Res. Cell Motil. 25, 645–655
- 78 Meriane, M., Roux, P., Primig, M., Fort, P. and Gauthier-Rouviere, C. (2000) Critical activities of Rac1 and Cdc42Hs in skeletal myogenesis: antagonistic effects of JNK and p38 pathways. Mol. Biol. Cell 11, 2513–2528
- 79 Bakiri, L., Lallemand, D., Bossy-Wetzel, E. and Yaniv, M. (2000) Cell cycle-dependent variations in c-Jun and JunB phosphorylation: a role in the control of cyclin D1 expression. EMBO J. 19, 2056–2068
- 80 Perdiguero, E., Ruiz-Bonilla, V., Serrano, A. L. and Munoz-Canoves, P. (2007) Genetic deficiency of p38α reveals its critical role in myoblast cell cycle exit: the p38α-JNK connection. Cell Cycle 6, 1298–1303
- 81 Trouillas, M., Saucourt, C., Duval, D., Gauthereau, X., Thibault, C., Dembele, D., Feraud, O., Menager, J., Rallu, M., Pradier, L. et al. (2008) Bcl2, a transcriptional target of p38α, is critical for neuronal commitment of mouse embryonic stem cells. Cell Death Differ. 15, 1450–1459
- 82 Wu, J., Kubota, J., Hirayama, J., Nagai, Y., Nishina, S., Yokoi, T., Asaoka, Y., Seo, J., Shimizu, N., Kajiho, H. et al. (2010) p38 mitogen-activated protein kinase controls a switch between cardiomyocyte and neuronal commitment of murine embryonic stem cells by activating myocyte enhancer factor 2C-dependent bone morphogenetic protein 2 transcription. Stem Cells Dev. 19, 1723–1734
- 83 Chai, Z., Yang, L., Yu, B., He, Q., Li, W. I., Zhou, R., Zhang, T., Zheng, X. and Xie, J. (2009) p38 mitogen-activated protein kinase-dependent regulation of SRC-3 and involvement in retinoic acid receptor α signaling in embryonic cortical neurons. IUBMB Life 61 670–678
- 84 Strassburger, M., Braun, H. and Reymann, K. G. (2008) Anti-inflammatory treatment with the p38 mitogen-activated protein kinase inhibitor SB239063 is neuroprotective, decreases the number of activated microglia and facilitates neurogenesis in oxygen-glucose-deprived hippocampal slice cultures. Eur. J. Pharmacol. 592, 55–61
- 85 Zhang, D., Guo, M., Zhang, W. and Lu, X. Y. (2011) Adiponectin stimulates proliferation of adult hippocampal neural stem/progenitor cells through activation of p38 mitogen-activated protein kinase (p38MAPK)/glycogen synthase kinase 3β (GSK-3β)/β-catenin signaling cascade. J. Biol. Chem. 286, 44913–44920
- 86 Oh, J. E., Bae, G. U., Yang, Y. J., Yi, M. J., Lee, H. J., Kim, B. G., Krauss, R. S. and Kang, J. S. (2009) Cdo promotes neuronal differentiation via activation of the p38 mitogen-activated protein kinase pathway. FASEB J. 23, 2088–2099
- 87 Chew, L. J., Coley, W., Cheng, Y. and Gallo, V. (2010) Mechanisms of regulation of oligodendrocyte development by p38 mitogen-activated protein kinase. J. Neurosci. 30, 11011–11027
- 88 Okamoto, S., Krainc, D., Sherman, K. and Lipton, S. A. (2000) Antiapoptotic role of the p38 mitogen-activated protein kinase-myocyte enhancer factor 2 transcription factor pathway during neuronal differentiation. Proc. Natl. Acad. Sci. U.S.A. 97, 7561–7566
- 89 Walshe, T. E., Leach, L. L. and D'Amore, P. A. (2011) TGF- β signaling is required for maintenance of retinal ganglion cell differentiation and survival. Neuroscience **189**, 123–131
- 90 Murry, T., Tabaee, A., Owczarzak, V. and Aviv, J. E. (2006) Respiratory retraining therapy and management of laryngopharyngeal reflux in the treatment of patients with cough and paradoxical vocal fold movement disorder. Ann. Otol. Rhinol. Laryngol. 115, 754–758
- 91 Poss, K. D., Wilson, L. G. and Keating, M. T. (2002) Heart regeneration in zebrafish. Science 298, 2188–2190
- 92 Raya, A., Koth, C. M., Buscher, D., Kawakami, Y., Itoh, T., Raya, R. M., Sternik, G., Tsai, H. J., Rodriguez-Esteban, C. and Izpisua-Belmonte, J. C. (2003) Activation of Notch signaling pathway precedes heart regeneration in zebrafish. Proc. Natl. Acad. Sci. U.S.A. 100 (Suppl. 1), 11889–11895
- 93 Nadal-Ginard, B. (1978) Commitment, fusion and biochemical differentiation of a myogenic cell line in the absence of DNA synthesis. Cell 15, 855–864
- 94 Tam, S. K., Gu, W., Mahdavi, V. and Nadal-Ginard, B. (1995) Cardiac myocyte terminal differentiation. Potential for cardiac regeneration. Ann. NY Acad. Sci. 752, 72–79
- 95 Beltrami, A. P., Barlucchi, L., Torella, D., Baker, M., Limana, F., Chimenti, S., Kasahara, H., Rota, M., Musso, E., Urbanek, K. et al. (2003) Adult cardiac stem cells are multipotent and support myocardial regeneration. Cell 114, 763–776
- 96 Oh, H., Bradfute, S. B., Gallardo, T. D., Nakamura, T., Gaussin, V., Mishina, Y., Pocius, J., Michael, L. H., Behringer, R. R., Garry, D. J. et al. (2003) Cardiac progenitor cells from adult myocardium: homing, differentiation, and fusion after infarction. Proc. Natl. Acad. Sci. U.S.A. 100, 12313–12318
- 97 Martin, R. L., McDermott, J. S., Salmen, H. J., Palmatier, J., Cox, B. F. and Gintant, G. A. (2004) The utility of hERG and repolarization assays in evaluating delayed cardiac repolarization: influence of multi-channel block. J. Cardiovasc. Pharmacol. 43, 369–379

- 98 Matsuura, K., Nagai, T., Nishigaki, N., Oyama, T., Nishi, J., Wada, H., Sano, M., Toko, H., Akazawa, H., Sato, T. et al. (2004) Adult cardiac Sca-1-positive cells differentiate into beating cardiomyocytes. J. Biol. Chem. 279, 11384–11391
- 99 Messina, E., De Angelis, L., Frati, G., Morrone, S., Chimenti, S., Fiordaliso, F., Salio, M., Battaglia, M., Latronico, M. V., Coletta, M. et al. (2004) Isolation and expansion of adult cardiac stem cells from human and murine heart. Circ. Res. 95, 911–921
- 100 Laugwitz, K. L., Moretti, A., Lam, J., Gruber, P., Chen, Y., Woodard, S., Lin, L. Z., Cai, C. L., Lu, M. M., Reth, M. et al. (2005) Postnatal isl1 + cardioblasts enter fully differentiated cardiomyocyte lineages. Nature 433, 647–653
- 101 Leri, A., Kajstura, J. and Anversa, P. (2005) Cardiac stem cells and mechanisms of myocardial regeneration. Physiol. Rev. 85, 1373–1416
- 102 Eriksson, M. and Leppa, S. (2002) Mitogen-activated protein kinases and activator protein 1 are required for proliferation and cardiomyocyte differentiation of P19 embryonal carcinoma cells. J. Biol. Chem. 277, 15992–16001
- 103 Tenhunen, O., Sarman, B., Kerkela, R., Szokodi, I., Papp, L., Toth, M. and Ruskoaho, H. (2004) Mitogen-activated protein kinases p38 and ERK 1/2 mediate the wall stress-induced activation of GATA-4 binding in adult heart. J. Biol. Chem. 279, 24852–24860
- 104 Han, J. and Molkentin, J. D. (2000) Regulation of MEF2 by p38 MAPK and its implication in cardiomyocyte biology. Trends Cardiovasc. Med. 10, 19–22
- Heidenreich, O., Neininger, A., Schratt, G., Zinck, R., Cahill, M. A., Engel, K., Kotlyarov, A., Kraft, R., Kostka, S., Gaestel, M. et al. (1999) MAPKAP kinase 2 phosphorylates serum response factor in vitro and in vivo. J. Biol. Chem. 274, 14434–14443
- 106 Ambrosino, C., Iwata, T., Scafoglio, C., Mallardo, M., Klein, R. and Nebreda, A. R. (2006) TEF-1 and C/EBP β are major p38 α MAPK-regulated transcription factors in proliferating cardiomyocytes. Biochem. J. **396**, 163–172
- 107 Chang, J., Sonoyama, W., Wang, Z., Jin, Q., Zhang, C., Krebsbach, P. H., Giannobile, W., Shi, S. and Wang, C. Y. (2007) Noncanonical Wnt-4 signaling enhances bone regeneration of mesenchymal stem cells in craniofacial defects through activation of p38 MAPK. J. Biol. Chem. 282, 30938–30948
- 108 Li, J., Zhao, Z., Yang, J., Liu, J., Wang, J., Li, X. and Liu, Y. (2009) p38 MAPK mediated in compressive stress-induced chondrogenesis of rat bone marrow MSCs in 3D alginate scaffolds. J. Cell. Physiol. 221, 609–617
- 109 Kawaki, H., Kubota, S., Suzuki, A., Suzuki, M., Kohsaka, K., Hoshi, K., Fujii, T., Lazar, N., Ohgawara, T., Maeda, T. et al. (2011) Differential roles of CCN family proteins during osteoblast differentiation: involvement of Smad and MAPK signaling pathways. Bone 49, 975–989
- 110 Greenblatt, M. B., Shim, J. H., Zou, W., Sitara, D., Schweitzer, M., Hu, D., Lotinun, S., Sano, Y., Baron, R., Park, J. M. et al. (2010) The p38 MAPK pathway is essential for skeletogenesis and bone homeostasis in mice. J. Clin. Invest. 120, 2457–2473
- 111 Nakajima, A., Sanjay, A., Chiusaroli, R., Adapala, N. S., Neff, L., Itzsteink, C., Horne, W. C. and Baron, R. (2009) Loss of Cbl-b increases osteoclast bone-resorbing activity and induces osteopenia. J. Bone Miner. Res. 24, 1162–1172
- 112 Maekawa, T., Jin, W. and Ishii, S. (2010) The role of ATF-2 family transcription factors in adipocyte differentiation: antiobesity effects of p38 inhibitors. Mol. Cell. Biol. 30, 613–625
- 113 Bhandari, D. R., Seo, K. W., Roh, K. H., Jung, J. W., Kang, S. K. and Kang, K. S. (2010) REX-1 expression and p38 MAPK activation status can determine proliferation/differentiation fates in human mesenchymal stem cells. PLoS ONE 5, e10493
- 114 Sellayah, D., Bharaj, P. and Sikder, D. (2011) Orexin is required for brown adipose tissue development, differentiation, and function. Cell. Metab. 14, 478–490
- 115 Hanley, S. and Rosenberg, L. (2007) Transforming growth factor β is a critical regulator of adult human islet plasticity. Mol. Endocrinol. **21**, 1467–1477
- 116 Hamamoto, K., Yamada, S., Hara, A., Kodera, T., Seno, M. and Kojima, I. (2011) Extracellular matrix modulates insulin production during differentiation of AR42J cells: functional role of Pax6 transcription factor. J. Cell Biochem. 112, 318–329
- 117 Ogihara, T., Watada, H., Kanno, R., Ikeda, F., Nomiyama, T., Tanaka, Y., Nakao, A., German, M. S., Kojima, I. and Kawamori, R. (2003) p38 MAPK is involved in activin A-and hepatocyte growth factor-mediated expression of pro-endocrine gene neurogenin 3 in AR42J-B13 cells. J. Biol. Chem. 278, 21693–21700
- 118 Yuan, L., Yu, Y., Sanders, M. A., Majumdar, A. P. and Basson, M. D. (2010) Schlafen 3 induction by cyclic strain regulates intestinal epithelial differentiation. Am. J. Physiol. Gastrointest. Liver Physiol. 298, G994–G1003
- 119 Kim, M. J., Park, B. J., Kang, Y. S., Kim, H. J., Park, J. H., Kang, J. W., Lee, S. W., Han, J. M., Lee, H. W. and Kim, S. (2003) Downregulation of FUSE-binding protein and c-myc by tRNA synthetase cofactor p38 is required for lung cell differentiation. Nat. Genet. 34, 330–336
- 120 Land, S. C. and Darakhshan, F. (2004) Thymulin evokes IL-6-C/EBP β regenerative repair and TNF- α silencing during endotoxin exposure in fetal lung explants. Am. J. Physiol. Lung Cell. Mol. Physiol. **286**, L473–L487

- 121 Kolosova, I., Nethery, D. and Kern, J. A. (2011) Role of Smad2/3 and p38 MAP kinase in TGF-β1-induced epithelial-mesenchymal transition of pulmonary epithelial cells.
 J. Cell. Physiol. 226, 1248–1254
- 122 Dong, J., Huang, S., Caikovski, M., Ji, S., McGrath, A., Custorio, M. G., Creighton, C. J., Maliakkal, P., Bogoslovskaia, E., Du, Z. et al. (2011) ID4 regulates mammary gland development by suppressing p38MAPK activity. Development 138, 5247–5256
- 123 Hsu, S., Dickinson, D., Borke, J., Walsh, D. S., Wood, J., Qin, H., Winger, J., Pearl, H., Schuster, G. and Bollag, W. B. (2007) Green tea polyphenol induces caspase 14 in epidermal keratinocytes via MAPK pathways and reduces psoriasiform lesions in the flaky skin mouse model. Exp. Dermatol. 16, 678–684
- 124 Connelly, J. T., Mishra, A., Gautrot, J. E. and Watt, F. M. (2011) Shape-induced terminal differentiation of human epidermal stem cells requires p38 and is regulated by histone acetylation. PLoS ONE 6, e27259
- 125 Kumar, S., Boehm, J. and Lee, J. C. (2003) p38 MAP kinases: key signalling molecules as therapeutic targets for inflammatory diseases. Nat. Rev. Drug Discov. 2, 717–726
- 126 Johnson, G. V. and Bailey, C. D. (2003) The p38 MAP kinase signaling pathway in Alzheimer's disease. Exp. Neurol. 183, 263–268
- 127 Broom, O. J., Widjaya, B., Troelsen, J., Olsen, J. and Nielsen, O. H. (2009) Mitogen activated protein kinases: a role in inflammatory bowel disease? Clin. Exp. Immunol. 158, 272–280
- 128 McFarland, L. V. (2008) State-of-the-art of irritable bowel syndrome and inflammatory bowel disease research in 2008. World J. Gastroenterol. 14, 2625–2629
- 129 Baumgart, D. C. and Carding, S. R. (2007) Inflammatory bowel disease: cause and immunobiology. Lancet 369, 1627–1640
- 130 Bassi, R., Heads, R., Marber, M. S. and Clark, J. E. (2008) Targeting p38-MAPK in the ischaemic heart: kill or cure? Curr. Opin. Pharmacol. 8, 141–146
- 131 Chung, K. F. and Adcock, I. M. (2008) Multifaceted mechanisms in COPD: inflammation, immunity, and tissue repair and destruction. Eur. Respir. J. 31, 1334–1356
- 132 Coulthard, L. R., White, D. E., Jones, D. L., McDermott, M. F. and Burchill, S. A. (2009) p38^{MAPK}: stress responses from molecular mechanisms to therapeutics. Trends Mol. Med. 15, 369–379
- 133 Cuenda, A. and Rousseau, S. (2007) p38 MAP-kinases pathway regulation, function and role in human diseases. Biochim. Biophys. Acta 1773, 1358–1375
- Mbalaviele, G., Anderson, G., Jones, A., De Ciechi, P., Settle, S., Mnich, S., Thiede, M., Abu-Amer, Y., Portanova, J. and Monahan, J. (2006) Inhibition of p38 mitogen-activated protein kinase prevents inflammatory bone destruction. J. Pharmacol. Exp. Ther. 317, 1044, 1062
- Ridley, S. H., Sarsfield, S. J., Lee, J. C., Bigg, H. F., Cawston, T. E., Taylor, D. J., DeWitt, D. L. and Saklatvala, J. (1997) Actions of IL-1 are selectively controlled by p38 mitogen-activated protein kinase: regulation of prostaglandin H synthase-2, metalloproteinases, and IL-6 at different levels. J. Immunol. 158, 3165–3173
- 136 Murphy, L. O. and Blenis, J. (2006) MAPK signal specificity: the right place at the right time. Trends Biochem. Sci. 31, 268–275
- 137 Tobiume, K., Matsuzawa, A., Takahashi, T., Nishitoh, H., Morita, K., Takeda, K., Minowa, O., Miyazono, K., Noda, T. and Ichijo, H. (2001) ASK1 is required for sustained activations of JNK/p38 MAP kinases and apoptosis. EMBO Rep. 2, 222–228
- 138 Roulston, A., Reinhard, C., Amiri, P. and Williams, L. T. (1998) Early activation of c-Jun N-terminal kinase and p38 kinase regulate cell survival in response to tumor necrosis factor α . J. Biol. Chem. **273**, 10232–10239
- 139 Okuma-Yoshioka, C., Seto, H., Kadono, Y., Hikita, A., Oshima, Y., Kurosawa, H., Nakamura, K. and Tanaka, S. (2008) Tumor necrosis factor-α inhibits chondrogenic differentiation of synovial fibroblasts through p38 mitogen activating protein kinase pathways. Mod. Rheumatol. 18, 366–378
- 140 Zwerina, J., Hayer, S., Redlich, K., Bobacz, K., Kollias, G., Smolen, J. S. and Schett, G. (2006) Activation of p38 MAPK is a key step in tumor necrosis factor-mediated inflammatory bone destruction. Arthritis Rheum. 54, 463–472
- 141 Ip, W. K., Wong, C. K. and Lam, C. W. (2006) Interleukin (IL)-4 and IL-13 up-regulate monocyte chemoattractant protein-1 expression in human bronchial epithelial cells: involvement of p38 mitogen-activated protein kinase, extracellular signal-regulated kinase 1/2 and Janus kinase-2 but not c-Jun NH2-terminal kinase 1/2 signalling pathways. Clin. Exp. Immunol. 145, 162–172
- 142 Reiner, S. L. (2007) Development in motion: helper T cells at work. Cell 129, 33–36
- 143 Danese, S., Semeraro, S., Marini, M., Roberto, I., Armuzzi, A., Papa, A. and Gasbarrini, A. (2005) Adhesion molecules in inflammatory bowel disease: therapeutic implications for gut inflammation. Dig. Liver Dis. 37, 811–818
- 144 Fiocchi, C. (1997) Intestinal inflammation: a complex interplay of immune and nonimmune cell interactions. Am. J. Physiol. 273, G769–G775
- 145 Scaldaferri, F., Sans, M., Vetrano, S., Correale, C., Arena, V., Pagano, N., Rando, G., Romeo, F., Potenza, A. E., Repici, A. et al. (2009) The role of MAPK in governing lymphocyte adhesion to and migration across the microvasculature in inflammatory bowel disease. Eur. J. Immunol. 39, 290–300

- 146 Ferrer, I., Blanco, R., Carmona, M., Puig, B., Barrachina, M., Gomez, C. and Ambrosio, S. (2001) Active, phosphorylation-dependent mitogen-activated protein kinase (MAPK/ERK), stress-activated protein kinase/c-Jun N-terminal kinase (SAPK/JNK), and p38 kinase expression in Parkinson's disease and dementia with Lewy bodies. J. Neural Transm. 108, 1383–1396
- 147 Culbert, A. A., Skaper, S. D., Howlett, D. R., Evans, N. A., Facci, L., Soden, P. E., Seymour, Z. M., Guillot, F., Gaestel, M. and Richardson, J. C. (2006) MAPK-activated protein kinase 2 deficiency in microglia inhibits pro-inflammatory mediator release and resultant neurotoxicity. Relevance to neuroinflammation in a transgenic mouse model of Alzheimer disease. J. Biol. Chem. 281, 23658–23667
- 148 Feijoo, C., Campbell, D. G., Jakes, R., Goedert, M. and Cuenda, A. (2005) Evidence that phosphorylation of the microtubule-associated protein Tau by SAPK4/p38δ at Thr50 promotes microtubule assembly. J. Cell Sci. 118, 397–408
- 149 Hanger, D. P., Anderton, B. H. and Noble, W. (2009) Tau phosphorylation: the therapeutic challenge for neurodegenerative disease. Trends Mol. Med. 15, 112–119
- Jin, S. X., Zhuang, Z. Y., Woolf, C. J. and Ji, R. R. (2003) p38 mitogen-activated protein kinase is activated after a spinal nerve ligation in spinal cord microglia and dorsal root ganglion neurons and contributes to the generation of neuropathic pain. J. Neurosci. 23, 4017–4022
- 151 Xu, J. T., Xin, W. J., Wei, X. H., Wu, C. Y., Ge, Y. X., Liu, Y. L., Zang, Y., Zhang, T., Li, Y. Y. and Liu, X. G. (2007) p38 activation in uninjured primary afferent neurons and in spinal microglia contributes to the development of neuropathic pain induced by selective motor fiber injury. Exp. Neurol. 204, 355–365
- 152 Mao, Z. and Wiedmann, M. (1999) Calcineurin enhances MEF2 DNA binding activity in calcium-dependent survival of cerebellar granule neurons. J. Biol. Chem. 274, 31102–31107
- 153 Muslin, A. J. (2008) MAPK signalling in cardiovascular health and disease: molecular mechanisms and therapeutic targets. Clin. Sci. 115, 203–218
- 154 Engel, F. B. (2005) Cardiomyocyte proliferation: a platform for mammalian cardiac repair. Cell Cycle 4, 1360–1363
- 155 Kaiser, R. A., Bueno, O. F., Lips, D. J., Doevendans, P. A., Jones, F., Kimball, T. F. and Molkentin, J. D. (2004) Targeted inhibition of p38 mitogen-activated protein kinase antagonizes cardiac injury and cell death following ischemia-reperfusion in vivo. J. Biol. Chem. 279, 15524–15530
- 156 Frantz, S., Behr, T., Hu, K., Fraccarollo, D., Strotmann, J., Goldberg, E., Ertl, G., Angermann, C. E. and Bauersachs, J. (2007) Role of p38 mitogen-activated protein kinase in cardiac remodelling. Br. J. Pharmacol. 150, 130–135
- 157 Streicher, J. M., Ren, S., Herschman, H. and Wang, Y. (2010) MAPK-activated protein kinase-2 in cardiac hypertrophy and cyclooxygenase-2 regulation in heart. Circ. Res. 106, 1434–1443
- 158 Zhou, L., Opalinska, J. and Verma, A. (2007) p38 MAP kinase regulates stem cell apoptosis in human hematopoietic failure. Cell Cycle 6, 534–537
- 159 Chang, Y. I., Hua, W. K., Yao, C. L., Hwang, S. M., Hung, Y. C., Kuan, C. J., Leou, J. S. and Lin, W. J. (2010) Protein-arginine methyltransferase 1 suppresses megakaryocytic differentiation via modulation of the p38 MAPK pathway in K562 cells. J. Biol. Chem. 285, 20595–20606
- 160 da Costa, S. V., Roela, R. A., Junqueira, M. S., Arantes, C. and Brentani, M. M. (2010) The role of p38 mitogen-activated protein kinase in serum-induced leukemia inhibitory factor secretion by bone marrow stromal cells from pediatric myelodysplastic syndromes. Leuk. Res. 34, 507–512
- 161 Fatrai, S., van Gosliga, D., Han, L., Daenen, S. M., Vellenga, E. and Schuringa, J. J. (2011) KRAS(G12V) enhances proliferation and initiates myelomonocytic differentiation in human stem/progenitor cells via intrinsic and extrinsic pathways. J. Biol. Chem. 286, 6061–6070
- Mayer, I. A., Verma, A., Grumbach, I. M., Uddin, S., Lekmine, F., Ravandi, F., Majchrzak, B., Fujita, S., Fish, E. N. and Platanias, L. C. (2001) The p38 MAPK pathway mediates the growth inhibitory effects of interferon-α in BCR-ABL-expressing cells. J. Biol. Chem. 276, 28570–28577
- 163 Bulavin, D. V. and Fornace, Jr, A. J. (2004) p38 MAP kinase's emerging role as a tumor suppressor. Adv. Cancer Res. 92, 95–118
- 164 Han, J. and Sun, P. (2007) The pathways to tumor suppression via route p38. Trends Biochem. Sci. 32, 364–371
- Haq, R., Brenton, J. D., Takahashi, M., Finan, D., Finkielsztein, A., Damaraju, S., Rottapel, R. and Zanke, B. (2002) Constitutive p38HOG mitogen-activated protein kinase activation induces permanent cell cycle arrest and senescence. Cancer Res. 62, 5076–5082
- Wang, W., Chen, J. X., Liao, R., Deng, Q., Zhou, J. J., Huang, S. and Sun, P. (2002) Sequential activation of the MEK-extracellular signal-regulated kinase and MKK3/6-p38 mitogen-activated protein kinase pathways mediates oncogenic Ras-induced premature senescence. Mol. Cell. Biol. 22, 3389–3403
- 167 Puri, P. L., Wu, Z., Zhang, P., Wood, L. D., Bhakta, K. S., Han, J., Feramisco, J. R., Karin, M. and Wang, J. Y. (2000) Induction of terminal differentiation by constitutive activation of p38 MAP kinase in human rhabdomyosarcoma cells. Genes Dev. 14, 574–584

168 Rossi, S., Stoppani, E., Puri, P. L. and Fanzani, A. (2011) Differentiation of human rhabdomyosarcoma RD cells is regulated by reciprocal, functional interactions between myostatin, p38 and extracellular regulated kinase signalling pathways. Eur. J. Cancer 47, 1095–1105

Received 6 March 2012/16 April 2012; accepted 18 April 2012 Published on the Internet 15 June 2012, doi:10.1042/BJ20120401 169 Noel, J. K., Crean, S., Claflin, J. E., Ranganathan, G., Linz, H. and Lahn, M. (2008) Systematic review to establish the safety profiles for direct and indirect inhibitors of p38 mitogen-activated protein kinases for treatment of cancer. A systematic review of the literature. Med. Oncol. 25, 323–330