FtsZ is a key protein involved in bacterial and organellar division. Bacteria have only one ftsZ gene, while chlorophytes (higher plants and green alga) have two distinct FtsZ gene families, named FtsZ1 and FtsZ2. This raises the question of why chloroplasts in these organisms need distinct FtsZ proteins to divide. In order to unravel new functions associated with FtsZ proteins, we have identified and characterized an Arabidopsis thaliana FtsZ1 loss-of-function mutant. ftsZ1-knockout mutants are impeded in chloroplast division, and division is restored when FtsZ1 is expressed at a low level. FtsZ1-overexpressing plants show a drastic inhibition of chloroplast division. Chloroplast morphology is altered in ftsZ1, with chloroplasts having abnormalities in the thylakoid membrane network. Overexpression of FtsZ1 also induced defects in thylakoid organization with an increased network of twisting thylakoids and larger grana. We show that FtsZ1, in addition to being present in the stroma, is tightly associated with the thylakoid fraction. This association is developmentally regulated since FtsZ1 is found in the thylakoid fraction of young developing plant leaves but not in mature and old plant leaves. Our results suggest that plastid division protein FtsZ1 may have a function during leaf development in thylakoid organization, thus highlighting new functions for green plastid FtsZ.

Key words: Arabidopsis thaliana, chloroplast division, FtsZ gene family, plastid division, plastid localization, thylakoid membrane.

INTRODUCTION

FtsZ plays an essential role in bacterial and archaeal division [1–3], in chloroplast division [4–6] and, in some lower organisms (algae and slime moulds), in mitochondrial division [4,7]. FtsZ, the most conserved of the bacterial cell division proteins, shares structural homology with the eukaryotic tubulin element and may be a progenitor of tubulin [8]. Bacterial FtsZ polymerizes like eukaryotic cytoskeletal proteins and forms the so-called Z ring, which forms a scaffold for assembly of at least ten other division proteins [9]. The Z ring adheres to the inside of the bacterial membrane and constricts to form a preseptal ingrowth, which then invaginates further to mediate division. Proteins involved in the cell division complex or divisome are cytoplasmic, periplasmic and membrane-embedded proteins. No outer membrane protein has so far been linked to cell division.

While most bacteria (including cyanobacteria) have only one FtsZ protein, plastid FtsZ protein sequences form three clades, including green-plastid FtsZ1, green-plastid FtsZ2 and the red and chromophyte algal group [10,11]. Phylogenetic analysis suggests that ftsZ gene duplication occurred subsequent to the endosymbiotic event. In chlorophytes, the gene duplication giving rise to the FtsZ1 and FtsZ2 gene families occurred before green algae branched from the ancestor of land plants [10–12]. The duplication event raises the question of why plastids in these organisms need two different proteins in order to divide when the ancestral bacteria use only one protein.

The major difference between FtsZ1 and FtsZ2 in both chlorophytes and non-chlorophytes lies in their C-terminal sequences. The C-terminal conserved motif, present in bacterial FtsZ, is required for direct interaction with ZipA and FtsA in Escherichia coli [13,14]. This domain is present in FtsZ2, but not in FtsZ1, in chlorophytes and in related sequences in non-chlorophytes. Homologues of ZipA and FtsA have not been identified in eukaryotic genomes, but the protein ARC6 (accumulation and replication of chloroplasts 6) in Arabidopsis shares homology with the cyanobacterial protein Ftn2 [15] and interacts with FtsZ2 via the C-terminal core domain [16], providing in planta evidence for a functional difference between the two FtsZ protein families in plants. The conserved N-terminal sequence is sufficient for bacterial FtsZ polymerization [17]. The major difference between FtsZ1 and FtsZ2 in higher plants concerns a single amino acid change in the conserved ‘tubulin signature motif’ [18].

In addition, FtsZ1 and FtsZ2 differ in their biochemical properties and subplastidial localization [19]. Expression of FtsZ1 and FtsZ2 in E. coli differentially affects division, and these different effects are related to the FtsZ2 C-terminal sequence. Only FtsZ1 is able to polymerize in vitro and forms GTP-dependent rod-shaped polymers and rings similar to the bacterial structures, but FtsZ2 can promote GTP-independent FtsZ1 polymerization. These results, together with other results showing the interaction of only FtsZ2 with ARC6 and an earlier expression of FtsZ2 during the cell cycle in BY2 cells [18], suggest that FtsZ2 and FtsZ1 fulfill different functions during chloroplast division [19].

In addition to being a key element in the chloroplast division machinery, plant FtsZ proteins may be involved either directly or indirectly in the co-ordination of cell division and plastid division.
An FtsZ1 isoform in the moss Physcomitrella patens is localized both in chloroplasts and in the cytoplasm, assembling into rings in both cell compartments, and transfected cells expressing high amounts of the protein were impeded in cell division [20]. In higher plants, expression of both FtsZ1 and FtsZ2 genes seems to be cell-cycle-regulated [18], and in Arabidopsis, the pre-replication factor ArCDT1 is involved not only in nuclear DNA replication but also in plastid division by means of an interaction with ARC6 [21].

Here, we describe the characterization of an Arabidopsis ftsZ1 mutant in which chloroplast division is strongly impeded but restored when FtsZ1 is expressed at a low level. FtsZ1 overexpression in both backgroungs [ftsZ1 and WT (wild-type)] induces a strong phenotype with a drastic effect on chloroplast morphology and division. Both null and FtsZ1-overexpressing plants have accelerated development. ftsZ1-deficient plants have an altered thylakoid membrane network, whereas FtsZ1 overexpression results in an increased network of twisting thylakoids and larger grana. We show that FtsZ1, in addition to being present in the stroma, is also found tightly associated with the thylakoid membranes and that this association is developmentally regulated.

**EXPERIMENTAL**

**Plant material, transformation and growth**

Arabidopsis, Wassilewskija ecotype (Ws), was used as WT plants for all experiments. The FtsZ1 T-DNA (transfer DNA) insertion mutant FST (flanking sequence tag) number 128A08 was obtained from the FLAdb/FST initiative (http://193.51.165.9/projects/FLAdb+/+HTML/index.shtml). Genomic DNA was amplified with the Tag6 (5′-CTTTTCATCTACGGGACATGCGACG-3′) and Tag7 (5′-GTCGATAAGAAAGGCAATTGTAG-3′) specific primers to detect the T-DNA. To follow the presence of the endogenous copy of the FtsZ1 gene and identify homozygous lines, genomic DNA was PCR-amplified with specific couples of primers: FtsZ1/D-FtsZ1/R and FtsZ1/D-Tag6 (FtsZ1/D: 5′-ATGGCGATAATTTCCGTGACAC-3′; FtsZ1/R: 5′-AGGGGCA-TCTGAAAAGAAGA-3′). Homozygous lines were propagated by repeated self-pollination and the absence of FtsZ1 expression was assayed by immunoblotting with polyclonal anti-FtsZ1 antibodies [18]. In order to exclude the presence of more than one T-DNA, Southern-blot analysis was performed. Nuclear DNA of the selected lines was digested with EcoRI, a restriction enzyme with a unique site in the FtsZ1 gene and identify homozygous lines, genomic DNA was PCR-amplified with specific couples of primers: FtsZ1/D-FtsZ1/R and FtsZ1/D-Tag6 (FtsZ1/D: 5′-ATGGCGATAATTTCCGTGACAC-3′; FtsZ1/R: 5′-AGGGGCA-TCTGAAAAGAAGA-3′). Homozygous lines were propagated by repeated self-pollination and the absence of FtsZ1 expression was assayed by immunoblotting with polyclonal anti-FtsZ1 antibodies [18]. In order to exclude the presence of more than one T-DNA, Southern-blot analysis was performed. Nuclear DNA of the selected lines was digested with EcoRI, a restriction enzyme with a unique site in the FtsZ1 gene and identify homozygous lines.
PBS) overnight at 4°C. Samples were then incubated with goat anti-rabbit IgG conjugated to 10 nm gold particles at a dilution of 1:20 in 1% BSA in PBS for 1 h at room temperature. Controls were performed by excluding the primary antibody. Finally, sections were post-stained with 5% uranyl acetate in water and lead citrate.

RESULTS
Phenotypic characterization of ftsZ1-deficient and FtsZ1-overexpressing plants

The Arabidopsis genome encodes a single FtsZ1 plastid division protein on chromosome 5 at the locus At5g55280. One Arabidopsis line containing a T-DNA insertion in exon 4 was identified by the Genoplante FLAGdb/FST initiative (Figure 1A). Plants homozygous for the mutant alleles were identified by PCR analysis of segregating plants (Figures 1B and 1C). Segregation values observed for kanamycin resistance (three-quarters resistant and one-quarter sensitive individuals after self-pollination of heterozygous plants) indicated that the mutant lines contained only one insertion of the T-DNA in their nuclear genome. The presence of only one T-DNA was also confirmed by Southern-blot analysis (Figure 1D). Only the 4.8 kbp EcoRI fragment, expected from the restriction data, was detected. Western-blot analysis confirmed the absence of FtsZ1 protein in the selected mutant lines (Figure 1E). Western blots showed FtsZ1 protein expression in WT plants and in the heterozygous line 1, but no expression in the homozygous lines 2, 3 and 4.

Mature ftsZ1 mesophyll and stem cells contained fewer, but larger chloroplasts than the corresponding cells in WT, although the cell sizes between the two plants were similar (Figures 2A, top panels, and 2B). The average chloroplast number at the equatorial plane decreased from 16 in WT mesophyll cells and 14 in WT stem cells to 7 and 4 respectively in ftsZ1 cells. The mutant phenotype was rescued by low-level expression of the FtsZ1 cDNA as shown by Western blotting, which did not alter the WT phenotype when expressed in WT plants (Figure 2A, middle panels). However, overexpression of FtsZ1 had a drastic effect on chloroplast division in both the WT and ftsZ1 backgrounds, with cells harbouring only few enlarged chloroplasts (Figure 2A, bottom panels). This corroborates previous results showing altered stoichiometry of FtsZ affecting chloroplast division [27]. As illustrated with the stem cells (Figure 2B), all cells of a given tissue were strongly affected by the absence or overexpression of the FtsZ1 protein. In all the mutant cells examined, the shape of the large chloroplasts was irregular, having lost the characteristic globular structure as observed in the WT plants. Compared with WT plants, ftsZ1 and FtsZ1-overexpressing lines exhibited accelerated growth (Figure 2C) and began flowering approx. 1 week earlier, on average, than WT plants (2 weeks after germination for the mutant plants compared with 3 weeks for the WT plants). ftsZ1 and 35S::FtsZ1 plants flowered with the same number of rosette leaves (an
average of 8 leaves) when the WT plants flowered with an average of 14 larger rosette leaves. After 6 weeks of growth, the size of the mutant plants was greatly reduced compared with the WT plants. Both mutant lines were able to produce viable seeds. Photosynthesis was investigated using pulse-amplitude-modulated chlorophyll fluorescence analysis and pigment content on 3-week-old plantlets. The $F_v/F_m$ ratio was identical between the mutant plants and the WT plants (Figure 2D). This indicates that the PSII reaction centre was properly assembled and photochemically competent in both mutant plants. Equally, the amount of photosynthetic pigments was unaltered in both mutant plants (Figure 2E).

**Chloroplast morphology and ultrastructure is affected in ftsZ1 and 3SS::FtsZ1 plants**

Three-week-old rosette leaves were fixed for TEM and observed at different levels of magnification. In WT plants, individual ellipsoidal chloroplasts, aligned along the cytoplasmic membrane, were observed at the cellular level (Figure 3A). These chloroplasts contained starch granules, and their internal structures, including evenly stacked granal thylakoids connected by stromal lamellae, were completely developed (Figures 3D and 3G). These lamellae are oriented along the convex side of the chloroplast facing the interior of the cell. In contrast, abnormally long chloroplasts were observed in the leaves of ftsZ1 (Figure 3B) and 3SS::FtsZ1 plants (Figure 3C). Although the long chloroplasts in the null plant were aligned along the plasma membrane, as was observed in the WT plant, the chloroplasts were detached from the plasma membrane at several points in the FtsZ1-overexpressing plant. Changes in thylakoid orientation were often observed in the null plant (Figure 3E; see also Supplementary Figure S1 at http://www.BiochemJ.org/bj/409/bj4090087add.htm), and membranes appeared less appressed (Figure 3H). Overexpression of FtsZ1 induced an increased network of twisting thylakoids (Figure 3F) and an increased number of thylakoids per grana stack (Figure 3I). An increase in starch granule number was observed in the null mutants, but no starch granules were observed in FtsZ1-overexpressing plants.
FtsZ1 is localized within the stroma, but is also found associated with the thylakoids

The changes in thylakoid organization in ftsZ1-null and ftsZ1-overexpressing plants indicated a possible role of FtsZ1 in thylakoid integrity and therefore suggested an association or at least some contact between FtsZ1 and thylakoid membranes. In order to test this hypothesis, we investigated the localization of FtsZ1 proteins in WT Arabidopsis plants by suborganellar fractionation and Western blotting, using an anti-FtsZ1 antibody previously shown to react with proteins from purified chloroplast extracts [28] (Figure 4A). The stromal, thylakoid and envelope fractions were purified from lysed chloroplasts by sucrose-density-gradient centrifugation [24]. Blots were first probed with the anti-FtsZ1 antibody and subsequently probed with antisera against (i) KARI (ketol-acid reducto-isomerase), a stromal protein involved in the amino acid biosynthetic pathway [29], (ii) PsbB (CP47), a thylakoid-integrated subunit of the PSII reaction centre, (iii) IEP37, an inner envelope protein of 37 kDa [30], and (iv) OEP21, an outer envelope protein of 21 kDa [31]. An FtsZ1 signal co-localized with the stromal and thylakoid controls, but not with the envelope proteins, suggesting that FtsZ1 localizes to thylakoid membranes (Figure 4A). Purified thylakoids were next washed with 1 M NaCl, 0.1 M Na$_2$CO$_3$ (pH 11.5) or 0.1 M NaOH to further characterize the association of FtsZ1 with the thylakoid membranes (Figure 4B). FtsZ1 was not released from the thylakoids by ionic extraction but was solubilized by alkaline extractions, indicating that it is peripherally associated with the membrane. The localization with the thylakoid membranes is independent of magnesium concentration because in the presence of EDTA, FtsZ1 is still detected in the thylakoid fraction (Figure 4B). As a control, release of PsbB from membranes was tested under identical conditions, but none was observed. It is therefore possible that FtsZ1 is associated with one leaflet of the membrane via lipid interaction.

To determine FtsZ1 topology, thermolysin proteolysis was performed with isolated thylakoids (Figure 4C). Under such a treatment, proteins on the stromal side of the thylakoid membrane are degradable, while lumen proteins, not accessible to thermolysin, are protected. FtsZ1 was accessible to thermolysin in the absence of EGTA, while the addition of EGTA inhibited the cleavage reaction, suggesting that FtsZ1 is located on the stromal-facing side of the thylakoid membranes.

To confirm the localization of FtsZ1 visually, immunoelectron microscopy was performed with 3-week-old Arabidopsis WT and FtsZ1-overexpressing plants (35S::FtsZ1). Small clusters and large clusters of gold particles in the WT and 35S::FtsZ1 plants respectively were observed in close association with the thylakoid
FtsZ1 was reported previously to be a soluble protein found only in the stromal fraction [19,32], while our results suggest that FtsZ1 is also associated with thylakoid membranes. To understand this discrepancy, we investigated whether FtsZ1 association with the thylakoids differed at various points during plant development. To test this hypothesis, we examined developmentally associated changes in FtsZ1 protein amount and localization by Western blotting. Proteins from subplastidial fractions were obtained from plants grown for 22, 35 and 43 days. A strong FtsZ1 signal was observed in the thylakoid fraction as well as in the stromal fraction of plants grown for 22 days (Figure 6). After 35 days of growth, only a faint signal was observed in the thylakoid fraction, while a strong signal was observed in the stromal fraction. FtsZ1 is no longer detected in the thylakoid fraction, but is still strongly present in the stroma in plants grown for 43 days. As expected, KARI was present only in the stroma, whereas PsbB was found exclusively in thylakoids, indicating the purity of the two fractions. The correlation between the level of FtsZ1 in the thylakoid fraction and the developmental stage of the tissue demonstrated the regulation of FtsZ1 localization within the chloroplast.

**DISCUSSION**

In order to highlight new plant-specific functions for FtsZ1, we have characterized an ftsZ1-null mutant of Arabidopsis. As expected from previous reports using an antisense strategy [27], ftsZ1 mutant cells contained fewer but larger chloroplasts than WT plants. Inhibition of chloroplast division is rescued by expressing FtsZ1 in these mutant plants, but the complementation is dose-dependent, since weak expression restores the WT phenotype, while a strong expression causes inhibition of chloroplast division and changes in chloroplast morphology, as previously shown [33]. Such a dose-dependent effect also occurs when FtsZ1 is expressed in *E. coli*, mimicking the expression of other bacterial ftsZ genes in *E. coli* [19]. These results confirm that a strict stoichiometry between subunits of the chloroplast division machinery is necessary for proper division. The overall life cycle of both ftsZ1 plants and FtsZ1-overexpressing lines is not affected, confirming that impaired plastid division has no significant effect on plant growth and development [34], although these plants are smaller than WT controls, flower earlier and contain a reduced number...
of rosette leaves. This phenotype is not a direct consequence of smaller cells and/or diminished photosynthetic yield, since the cell size and photosynthetic capacity, as determined by measuring the maximal photochemical efficiency of PSII and quantifying the photosynthetic pigments, are unaltered in both transgenic lines. Developmental variation in the rate of leaf initiation has been suggested to result from change in the rate of cell division in the SAM (shoot apical meristem) [35]. The phenotype observed in the mutant plants might be driven by changes in cell division in the SAM, thus connecting cell division and plastid division as suggested by a number of studies [18,20,21,36].

Mesophyll chloroplast ultrastructure is affected differently in the null and FtsZ1-overexpressing plants. FtsZ1 chloroplasts have frequent changes in their thylakoid orientation and fewer thylakoids per granal stack, while FtsZ1-overexpressing chloroplasts display a highly disturbed thylakoid network with an increase in granal stacking. Organizational changes of the thylakoid membranes within some, but not all, chloroplast division mutants have also been described. Granal stacks in GC1 (giant chloroplasts 1)-deficient giant chloroplasts are more densely packed than in WT plants [37]. Thylakoids in arc5 and arc6 chloroplasts are also highly stacked when compared with WT chloroplasts and show increased stacking when plants are grown under high light [38]. Under the same conditions, a decrease is observed with WT plants. Interestingly, FtsZ1-overexpressing plants show an increased network of twisting thylakoids similar to those observed in arc3 [38]. These results demonstrate that some proteins involved in plastid division also affect chloroplast internal structures. GC1 is plastid-localized and is anchored to the stromal surface of the chloroplast inner envelope [37]. Arc3 encodes a cytosolic dynamin-like protein and forms a cytosolic ring structure on the outside of the chloroplast [39]. Arc6, which encodes a J-domain protein and is a homologue of the cyanobacterial cell division protein Ftn2 [15], has been shown to be an integral inner envelope membrane protein. Arc3, a fusion of FtsZ and PIP5K (phosphatidylinositol-4-phosphate 5-kinase) [40], has recently been shown to be located in the stroma [41]. None of these proteins have been found associated with thylakoids, suggesting that the effect on the thylakoid network in these mutants is indirect. The twisting thylakoids observed in arc3, resembling those observed in the FtsZ1-overexpressing plants, might result from the misplacement of FtsZ1. As a matter of fact, Arc3 has been shown to interact specifically with FtsZ1, acting in division site placement [41].

The localization of FtsZ proteins in higher plants is more complex. Both FtsZ1 and FtsZ2 have been found in the chloroplast stromal compartment in Arabidopsis chloroplasts [32], but localization in the thylakoid fraction has not been addressed previously. We have shown that FtsZ1 and FtsZ2 are localized within the stroma in chloroplasts from mature spinach (Spinacia oleracea) leaves, but that FtsZ2 is also found associated with the chloroplast envelopes [19]. In the present study, we show that FtsZ1 is present in the stromal and thylakoid fractions from the chloroplasts of young Arabidopsis seedlings. Moreover, we demonstrate that the levels of FtsZ1 associated with the thylakoids decreases with the age of the plants. This result shows that FtsZ1 localization within the chloroplast is developmentally regulated and suggests that the protein fulfills specific functions at different stages of plant development. It is not known yet whether the difference in localization between spinach and Arabidopsis is a consequence of the different developmental stage and/or culture conditions of the analysed plants. In the previous experiments, spinach leaves, purchased from a local market, were grown under external conditions, while Arabidopsis plants in the present study were grown under strict controlled temperature and light conditions. It is not known so far whether FtsZ1 association with the thylakoids is dependent on these growth conditions. Alternatively, it is possible that the discrepancy reflects species difference.

The thylakoid organization phenotypes associated with the absence or overexpression of FtsZ1 in the mutant plants corroborate the hypothesis that FtsZ1 is involved in determining thylakoid morphology. It is therefore conceivable that the organizational changes of the thylakoid membranes within the arc chloroplasts reflect an increased localization of FtsZ1 within the thylakoids in the absence of the ARC proteins. Interestingly, dynamin-like proteins, which are involved in plastid division [39], have also been shown to be a determinant in thylakoid morphology [42,43]. In Arabidopsis, FZL, which is related to FZO, a dynamin protein involved in mitochondrial morphology in fungi and animals, is distributed between the thylakoid and envelope membranes. FZL mutants have abnormalities in the morphology and distribution of granal and stromal thylakoids [43]. The results presented in this paper suggest that the dynamic duo of FtsZ and dynamin, which has been shown to be involved in plastid division, participates in the remodelling of the thylakoid membranes. FtsZ1 and FZL are not involved in the early steps of thylakoid biogenesis since both ftsZ1 and fzo chloroplasts have a preformed thylakoid network. Other proteins such as VIPP1 (vesicle-inducing protein in plastids 1) [44] and Thf1 (thylakoid formation 1) [45] have been implicated in chloroplast vesicle trafficking and have been shown to affect thylakoid formation. The association of FtsZ1 with the thylakoids during leaf expansion suggests that FtsZ1 might be involved in thylakoid membrane partitioning to daughter plastids during division and in the re-arrangement of the internal membrane structures when new thylakoid membranes are synthesized, thus highlighting specific functions for green plastid FtsZ.

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